

# GENOTYPING BY PCR PROTOCOL

## MUTANT MOUSE RESOURCE & RESEARCH CENTER: UC DAVIS

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530-754-MMRRC

Protocol Name: B6:129S5-K<sup>tm1Lex</sup>/Mmucd MMRRC: 011732-UCD

**Protocol:**

Reagent/Constituent	Volume (μL)
Water	10.275
10x Buffer	2.5
MgCl <sub>2</sub> (stock concentration is 25mM)	1.7
Betaine (stock concentration is 5M) <i>Optional</i>	6.5
dNTPs (stock concentration is 10mM)	0.5
DMSO <i>Optional</i>	0.325
Primer 1. (stock concentration is 20μM)	0.5
Primer 2. (stock concentration is 20μM)	0.5
Primer 3. (stock concentration is 20μM)	0.5
Primer 4. (stock concentration is 20μM)	0.5
Taq Polymerase 5Units/μL	0.2
DNA (example) extracted w/ "Qiagen DNeasy columns or other similar silica based kits"	1.0
<b>TOTAL VOLUME</b>	<b>25</b>

**Comments on protocol:**

- Protocol may work with other DNA extraction methods.
- Use Touch-Down cycling protocol-first 10 cycles anneal at 65°C decreasing in temperature by 1.0°C; next 30 cycles anneal at 55°C.
- Betaine and DMSO have been standardized due to high GC content. Protocol may be tested without. Also, may adjust MgCl<sub>2</sub> to increase reaction or decrease non-specific amplifications.

**Strategy:**

Steps		Temp (°C)	Time (m:ss)	# of Cycles
1. Initiation/Melting	HOT START? <input type="checkbox"/>	94	5:00	1
2. Denaturation		94	0:15	
3. Annealing	steps 2-3-4 cycle in sequence	65 to 55 (↓1°C/cycle)	0:30	<b>40x</b>
4. Elongation		72	0:40	
5. Amplification		72	5:00	1
6. Finish		15	∞	n/a

**Primers:**

**Electrophoresis Protocol:**

Name	Nucleotide Sequence (5' - 3')	Agarose :1.5%	V:90
1. Primer 0787-12	GATGGGGTTCGACGTC	Running Time: 90 min.	
2. Primer 0787-13	TAAAGGAGGAAAGCCATTGTC	<b>Primer</b>	<b>Band (bp)</b>
3. Primer Neo3a	GCAGCGCATCGCCTTCTATC	1 & 2	186
4. Primer 0787-20	ATGCTCCAGACATTCTCAGC	3 & 4	455
			<b>Genotype</b>
			wildtype
			mutant

