

GENOTYPING BY PCR PROTOCOL FORM MUTANT MOUSE RESOURCE & RESEARCH CENTER

sacoord@mmrrc.org

800-910-2291 North America, +1-530-757-5710 International

Please provide the following information required for genetic analysis of your mutant mice.

Please fill in form electronically by tabbing through the text fields.

The first 2 pages are protected with gray text fields available as needed to describe your protocol. The last page is not protected to allow for pictures and or comments that will not fit on the protected form.

Thank you!

Note to MAC users: to ensure your graphic can be viewed on a PC please follow the steps below when inserting the graphic into this document. **DO NOT** drag and drop or copy/paste the graphic into this document.

- Open the original graphic in the program that created it
- Choose File, Save As
- Select No Compression in the save options.
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- Switch to Word, choose Insert, Picture, From File and choose the newly saved picture.

These instructions are very generic. The menu options for your graphics program may be different.

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Telephone (858) 822-2298	FAX	
Strain Name nur12 Chr17-129SvlmJ on BALB/c		MMRRC Stock Number 41446

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NAME OF PCR: _____ MMRRC # 041446-

Protocol: (PCR protocol provided by Donating Investigator)

Reagent/ Constituent	Volume (µL)
Water	7.61
10x Buffer (contains 15mM MgCl ₂)	2
dNTPs (stock concentration is 10mM)	0.36
Primer 1 (stock concentration is 20µM) forward	0.2
Primer 2 (stock concentration is 20µM) reverse	0.2
Primer 3 (stock concentration is µM)	
Primer 4 (stock concentration is µM)	
Taq Polymerase 5units/ul	0.18
Additives / Other (if applicable): BSA 10mg/ml	0.15
DNA sample extracted <input type="checkbox"/> NaOH <input checked="" type="checkbox"/> Proteinase K <input type="checkbox"/> Other:	5
TOTAL VOLUME OF REACTION:	5.330 µL

Comments on protocol (e.g., different concentration of MgCl₂, etc):

25mM MgCl₂

Strategy:

Steps	Temp (°C)	Time (m:ss)	# of Cycles
1. Initiation/Melting HOT START? <input checked="" type="checkbox"/>	95	3:00	1
2. Denaturation	95	0:15	} 35
3. Annealing } steps 2-3-4 will cycle in sequence	55	0:15	
4. Elongation	72	0:30	
5. Amplification (i.e., 72°C, 10 min)	72	6:00	1
6. Finish (i.e., 4°C, indefinite)	4	n/a	n/a

Primers:

Name	Nucleotide Sequence (5' - 3')
1: D15Mit226	ATCTCACGCTTCTCCCTTCA TGATGCTTGGATCTTGTGGA
2: D15Mit209	TTGTGCTTCACTAGATGTAGACCA TTTTATAGTTGCACATAAGCAGCA
3: D15Mit107	CAACACTTATACTTGTGTCAGGG TCATGGTTGGAACAGCAGAC
4: D15Mit245	#4: ACCAATACACTTCATGCAAACG CAGTGACCGTAGGTTCAATAACC
5: D15Mit161	#5: Primer 1 Sequence TCTGTTTTGTTTGTTCGTTTGC Primer 2 Sequence TAAAATCTCCCTGTATACAAGTCTGTG

Electrophoresis Protocol:

Agarose: 3%Superfine V: 275

Estimated Running Time: 40 min.

Primer Combination	Band	Genotype
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	bp	
	bp	
	bp	

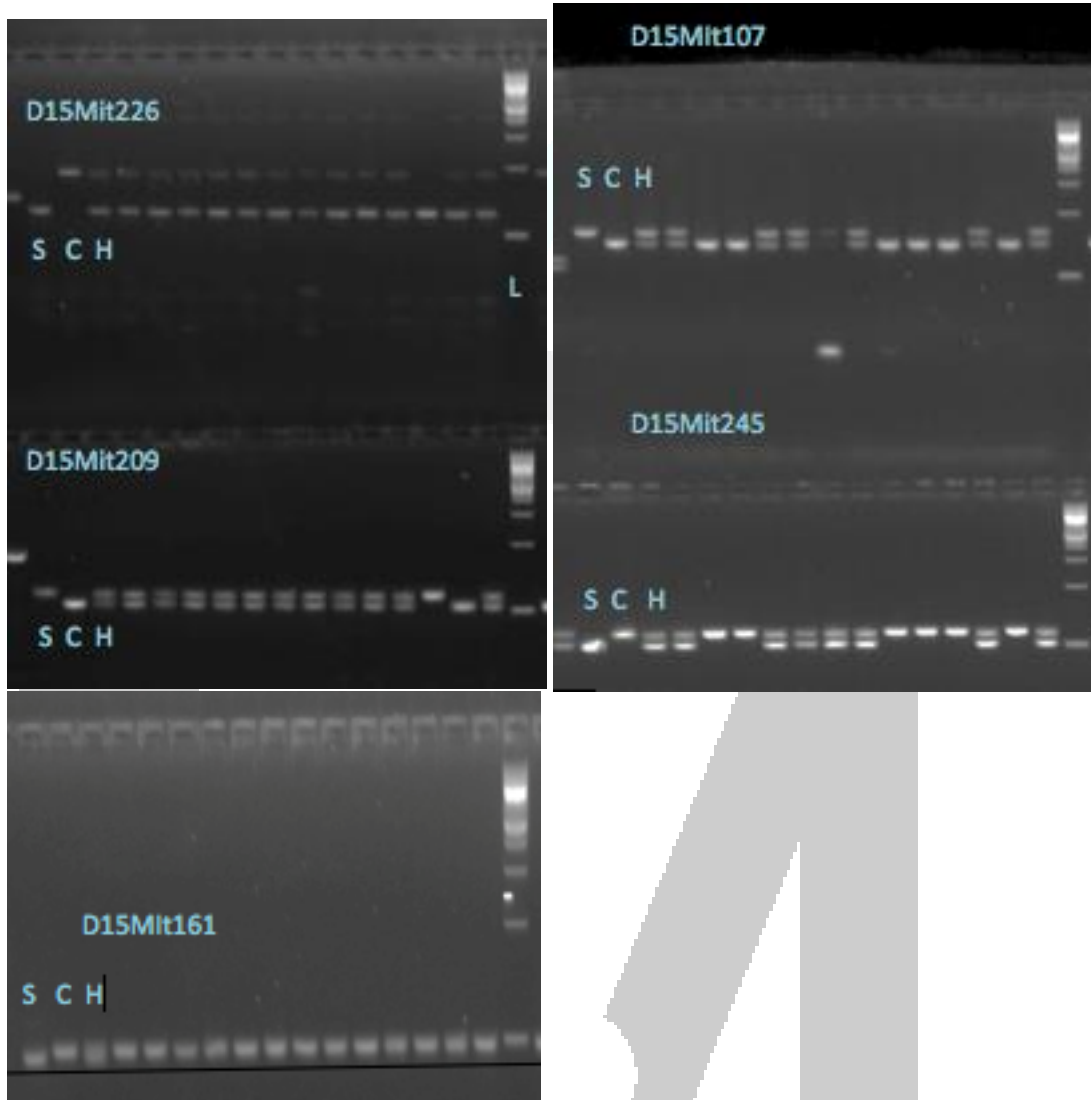


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Protocol / Gel Comments: Ladder= 100bp; S= 129Svlm/J; C=Balb/c; H= heterozygote



Lanes:
1. 1Kb+ ladder
2. H₂O
3. Wild-type +/- sample
4-5. Hom -/- samples

Sample Gel:

PCR protocol provided by Donating Investigator