

**GENOTYPING BY PCR PROTOCOL**  
**MUTANT MOUSE RESOURCE & RESEARCH CENTER: UC DAVIS**

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**Protocol Name:** CR1309 Fam19a3 EXDEL

**Protocol:**

Reagent/Constituent	Volume ( $\mu$ L)
Water	10.775
10x Buffer	2.5
MgCl <sub>2</sub> (stock concentration is 25mM)	1.7
Betaine (stock concentration is 5M) <i>Optional</i>	6.5
dNTPs (stock concentration is 10mM)	0.5
DMSO <i>Optional</i>	0.325
Primer 1. (stock concentration is 20 $\mu$ M) comF	0.6
Primer 2. (stock concentration is 20 $\mu$ M) wtR	1.2
Primer 3. (stock concentration is 20 $\mu$ M) mutR	0.3
Taq Polymerase 5Units/ $\mu$ L	0.2
DNA (example) extracted w/ "Qiagen DNeasy columns or other similar silica based kits"	1.0
<b>TOTAL VOLUME OF REACTION:</b>	<b>25.000 <math>\mu</math>L</b>

**Comments on protocol:**

- Protocol may work with other DNA extraction methods.
- Use Touch-Down cycling protocol-first 10 cycles anneal at 65°C decreasing in temperature by 1.0°C; next 30 cycles anneal at 55°C.
- Betaine and DMSO have been standardized due to high GC content. Protocol may be tested without. Also, may adjust MgCl<sub>2</sub> to increase reaction or decrease non-specific amplifications.

**Strategy:**

Steps	HOT START? <input type="checkbox"/>	Temp (°C )	Time (m:ss)	# of Cycles
1. Initiation/Melting		94	5:00	1
2. Denaturation		94	0:15	
3. Annealing	steps 2-3-4 cycle in sequence	65 to 55 ( $\downarrow 1^{\circ}\text{C}/\text{cycle}$ )	0:30	<b>40x</b>
4. Elongation		72	0:40	
5. Amplification		72	5:00	1
6. Finish		15	$\infty$	n/a

**Primers:**

Name	Nucleotide Sequence (5' - 3')	Agarose: 1.5% V: 90	Electrophoresis Protocol:	
1. CR_Fam19a3-comF	CTCCAACATCCGCAGGTCTGG	Estimated Running Time: 90 min.		
2. CR_Fam19a3-wtR	GACAGGCAGGAGCATTCGACC	Primer Combination	Band (bp)	Genotype
3. CR_Fam19a3-mutR	GCTTCTGTTAGTTAACCTGCTTGCTG	1 & 2, 1 & 3	749,2409*	wildtype
		1 & 3	500	mutant

**Allele Description:** Exon 5-6 ([ENSMUSE00000424938](#), [ENSMUSE00000424932](#)) and flanking splicing regions were constitutively deleted from the Fam19a3 gene [ENSMUST00000139783.1](#) using CRISPR Cas9 gene editing technology in mouse zygotes. Subsequent founders were backcrossed to C57BL6/N to produce sequence confirmed heterozygous animals.

\*May not see larger wildtype band

