

**GENOTYPING BY PCR PROTOCOL  
MUTANT MOUSE REGIONAL RESOURCE CENTER**

[sacoord@mmrrc.org](mailto:sacoord@mmrrc.org)

800-910-2291 North America, +1-530-757-5710 International

Please provide the following information required for genetic analysis of your mutant mice.

*Note to MAC users: to ensure your graphic can be viewed on a PC please follow the steps below when inserting the graphic into this document. DO NOT drag and drop or copy/paste the graphic into this document.*

- *Open the original graphic in the program that created it*
- *Choose File, Save As*
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*These instructions are very generic. The menu options for your graphics program may be different.*

|  |                                    |                     |
|--|------------------------------------|---------------------|
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| Telephone<br><b>858-534-7809</b>                           | FAX                                |                     |
| Strain Name<br><b>P53/Y240F</b>                            | MMRRC Stock Number<br><b>43554</b> |                     |

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NAME OF PCR: Y240F

MMRRC: 0-CTR

Protocol:

*(PCR protocol provided by Donating Investigator)*

| Reagent/Constituent   | Volume (μL)  |
|---|--------------|
| Water   | 28.5         |
| 10x Buffer  | 5            |
| MgCl <sub>2</sub> (stock concentration is mM)   |              |
| Betaine (stock concentration is 5M) <i>Optional</i>   |              |
| dNTPs (stock concentration is 10mM)   | 1            |
| DMSO <i>Optional</i>  |              |
| Primer 1. (stock concentration is 20μM)   | 5            |
| Primer 2. (stock concentration is 20μM)   | 5            |
| Primer 3. (stock concentration is 20μM)   |              |
| Primer 4. (stock concentration is 20μM)   |              |
| Taq Polymerase 5Units/μL  | .5           |
| DNA (50-200ng/ μL) extracted w/ "Qiagen DNeasy columns or other similar silica based kits"  | 5            |
| <i>The total volume is auto-calculated based on volumes entered, right click the total and update field to show/recalculate the total volume.</i> |              |
| <b>TOTAL VOLUME OF REACTION:</b>  | <b>50 μL</b> |

Comments on protocol:

Strategy:

| Steps  | Temp (°C ) | Time (m:ss) | # of Cycles |
|--|------------|-------------|-------------|
| 1. Initiation/Melting <span style="float:right">HOT START? <input type="checkbox"/></span> | 94         | 3           | 1           |
| 2. Denaturation  | 94         | 2           |             |
| 3. Annealing <span style="float:right">steps 2-3-4 cycle in sequence</span>                | 51         | :45         | 40x         |
| 4. Elongation  | 72         | 1           |             |
| 5. Amplification   | 72         | 5           | 1           |
| 6. Finish  | 4          | ∞           | n/a         |

Primers:

Electrophoresis Protocol:

| Name       | Nucleotide Sequence (5' - 3') | Argarose: 1.2%                   | V: 70    |
|------------|-------------------------------|----------------------------------|----------|
| 1. Exon 7F | CAG ATC CTC AGT TTG TGG TCT   | Estimated Running:Time: 120 min. |          |
| 2. Exon7R  | CAG GTG AGT CTG CTT ACA TG    | Primer Combination               | Band     |
| 3.         |                               | Exon7F + Exon7R                  | 350 bp   |
| 4.         |                               |                                  | bp       |
| 5.         |                               |                                  | bp       |
|            |                               | Genotype                         | Sequence |

*Please size gel images to fit in this space*

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**Protocol / Gel Comments:**

After running the PCR, purify, and send to sequencing with Exon7F and see if wildtype, homo, or het.  
For the P53 PCR protocol, please refer to The Jackson Lab stock#002101.

**Gel pictures:**



